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# PRODUCT-SPECIFIC PROTOCOLS

## IF (CL488-66071)

| Sample type | Fixation method | Blocking buffer | Antibody dilution buffer | Primary antibody dilution | Incubation time |
|---|---|---|---|---|---|
| HeLa cells | -20°C Ethanol, 10 min at room temp | 3% BSA in 1×PBS | 1% BSA in 1×PBS | 1:100 | 1.5 h at room temp |
| | Secondary antibody | | Secondary antibody dilution | Incubation time | Mounting media |
| | - | | | Hydromount? mounting media with DAPI | |

# PROTOCOL (more details available upon request):

1. Seed cells on sterile glass coverslips and gently wash the coverslip with 1X PBS.
2. Fix the samples with -20°C Ethanol, 10 min at room temp.
3. Rinse coverslip with 1XPBS 3 times for 5 min each.
4. Incubate the samples with 0.2% Triton X-100 made in 1XPBS for 5 min at room temperature.
5. Rinse the samples 3 times with 1XPBS for 3 min each.
6. Incubate the samples with Blocking buffer at room temperature for 1 h.
7. Aspirate Blocking buffer and incubate with primary antibody at room temperature for 2 hours or at 4°C for overnight.
8. Wash samples with 1XPBS 3 times for 5 min each.
9. Mount and seal the sample, then visualize the target antigen using a fluorescent microscope.

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